# Contributions to the lichen genus Crypthonia (Arthoniales) in India

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The lichen genus Crypthonia in India is represented by two species namely, Crypthonia albida (Fée) Frisch & G. Thor and Crypthonia palaeotropica Frisch & G. Thor. The taxonomic descriptions and diagnostic characteristics that distinguish these from allied species are discussed. Brief notes on geographic distribution, ecology, along with a world distribution map and key of all known species of Crypthonia are provided.

ABSTRACT

# **INTRODUCTION**

The genus Crypthonia Frisch & G. Thor (Arthoniaceae) was established by Frisch and Thor (2010) to accommodate species having Herpothallon-like thalli with maculate ascomata, asci, and ascospores similar to those found in Arthonia s. l. (Frisch et al. 2014). So far, the genus comprises 16 species globally (Frisch & Thor 2010; Menezes et al. 2013; Aptroot et al. 2014; Cáceres & Aptroot 2016; Sipman 2018). The genus is generally characterized by its loosely attached byssoid thallus with colored prothallus, ascomata lacking a distinct exciple, asci of the Arthonia-type, lacking a hemiamyloid ring in the tholus, transversely septate to submuriform, hyaline ascospores, lacking lichen substances or with mainly divaricatic, gyrophoric, norstictic, psoromic, usnic acids, and lichexanthone.

During the ongoing revisionary studies on the Arthoniales of India, some of the interesting specimens having byssoid thallus and colored hypothallus were studied in detail, which resulted in two species of Crypthonia. These specimens belonged to the Western Ghats, and hence the species recorded their extended distribution from the earlier known localities of Assam, Andaman Islands, and Darjeeling hills. A detailed taxonomic account of both the species and world key is provided.

# **MATERIALS AND METHODS**

The specimens examined in the present study were housed at herbarium LWG (CSIR-National

Botanical Research Institute, Lucknow). The morphological details were examined using a Leica S8APO stereomicroscope, and anatomical details were studied using a Leica DM2500 light microscope equipped with camera and image analysis software. The hand-cut sections of thalli and ascomata mounted in distilled water, KOH solution (K), and lactophenol cotton blue (LPCB) were studied. The amyloid reactions were tested in Lugol's iodine solution without (I) or with pretreatment with KOH (KI). All the measurements were made on material mounted in distilled water. The chemistry was studied by spot tests and thin layer chromatography (in solvent system C) following Orange et al. (2001). The world distribution map of Crypthonia is prepared by using the software QGIS 3.6 (Fig. 2). The GPS coordinates were procured from the published references (Table 1). The species for which geographical coordinates were not available, were georeferenced by use of specimens examined information and Google Earth Pro 7.3.

# THE SPECIES

1. Crypthonia albida (Fée) Frisch & G. Thor, Mycol. Progr. 9(2): 290. 2010. Hypochnus albidus Fée, Essai crypt. écorces exot. offic., suppl. et révis.: 13. 1837. Chiodecton sterile Müll. Arg., Mém. Soc. Phys. Hist. Nat. Genève 29(8): 65. 1887, non Chiodecton albidum (Taylor) Leight. Herpothallon albidum (Fée) Aptroot, Lücking & G. Thor, Biblioth. Lichenol. 99: 29. 2009. (Fig. 1A).

Thallus corticolous, up to 8 cm diam., greyish to pale yellowish green, byssoid, more or less loosely appressed to the substrate,  $120-250 \ \mu m$  thick, with scattered calcium oxalate crystals,  $2-10 \ \mu m$  in diam. Hypothallus dark olivaceous-green, byssoid, hyphae  $1-3 \ \mu m$  wide. Prothallus white, up to 3 mm wide, fibrous-

like. Pseudoisidioid outgrowths few to numerous, polymorphic, irregularly cushion-shaped, fluffy-felty with many projecting hyphae at the centre of the thallus, small globular to short branched at periphery regions, whitish, up to  $2 \times 1$  mm. Photobiont cells  $5-14 \times 4-9$  µm. Ascomata and pycnidia not seen.

*Chemistry*: Thallus K-, C-, P+ yellow, UV-; psoromic acid detected in TLC.

*Remarks: C. albida* is a sterile species having *close morphological* similarity to *C. palaeotropica* Frisch & G. Thor, which has an olive-brown to blackish rather than dark green hypothallus and more compact pseudoisidia (Frisch and Thor 2010). The species is somewhat similar in thallus structure to *Dichosporidium nigrocinctum* (Ehrenb.) G. Thor. The fluffy outgrowths (pseudoisidia) of *C. albida* 



Fig. 1: (A) habit of *Crypthonia albida*, (B–C) *C. palaeotropica*, (B) habit, (C) section of ascomata showing asci, ascospores and paraphysoids mounted in Lugol's iodine solution. Scale bar: (A–B) 1 cm, (C) 50 µm.



Fig. 2: Map of world distribution of *Crypthonia* species. (A) *C. albida*, (B) *C. athertoniensis*, (C) *C. bella*, (D) *C. biseptata*, (E) *C. brevispora*, (F) *C. citrina*, (G) *C. corticorygmoides*, (H) *C. divaricatica*, (I) *C. lichexanthonica*, (J) *C. mycelioides*, (K) *C. olivacea*, (L) *C. palaeotropica*, (M) *C. polillensis*, (N) *C. streimannii*, (O) *C. submuriformis*, (P) *C. vandenboomii*.

appear as reminiscent of the cylindrical pseudoisidia of *D. nigrocinctum* but remain short and irregular in structure (Aptroot et al. 2009). The species was first reported from India as *Herpothallon albidum* from West Bengal by Jagadeesh Ram & Sinha (2009). Later, they reidentified it as *Crypthonia palaeotropica* (Jagadeesh Ram & Sinha 2017). Behera et al. (2021) mentioned the occurrence of this species (as *H. albidum*) in Assam.

*Specimens examined*: India, Karnataka, Uttara Kannada district, Karwar-Hubli road, 24 February 2018, on bark, S. Nayaka & party 18-029164 (LWG); *ibid.*, Shimoga dist., Hosanagra, Ripponpete, elev. 693 m a.s.l., N13°55'44.9", E75°15'40.7", 21 February 2018, on bark, S. Nayaka & party 18-034281 (LWG); *ibid.*, Sagara taulk, Honnemaradu, elev. 643 m a.s.l., 19 March 2001, on the bark of *Olea dioica* Roxb., S. Nayaka 01-107278 (LWG); *ibid.*, Mattikoppa, elev. 603 m a.s.l., 17 March 2001, on bark, S. Nayaka 01-67381, 01-67386, 01-67392 (LWG); *ibid.*, Negiloni, elev. 710 m a.s.l., 18 March 2001, on the bark of *Aglaia elaegnoidea* 

(Juss.) Benth., S. Nayaka 01-107239 (LWG); *ibid.*, Holebagilu, elev. 623 m a.s.l., 16 March 2001, on the bark of *Diospyros crumenata* Thw., S. Nayaka 01-222800 (LWG); *ibid.*, Karumane, elev. 599 m a.s.l., 21 March 2001, on the bark of *Pterospermum reticulatum* Wight & Arn., S. Nayaka 01-66197 (LWG); *ibid.*, Hubse, elev. 594 m a.s.l., 16 March 2001, on the bark of *Ventilago maderaspatana* Gaertn., S. Nayaka 01-222736 (LWG); *ibid.*, on the bark of *Artocarpus integer* (Thunb.) Merr., S. Nayaka 01-222706 (LWG); *ibid.*, on the bark of *Ficus* sp., S. Nayaka 01-222728 (LWG).

**2.** *Crypthonia palaeotropica* Frisch & G. Thor, Mycol. Progr. 9(2): 300. 2010. (Fig. 1B&C).

Thallus corticolous, up to 10 cm diam., surface pale greenish, byssoid, more or less loosely attached to the substrate, 100–300  $\mu$ m thick, calcium oxalate crystals numerous, 5–12  $\mu$ m in diam. Hypothallus dark olivaceous brown, byssoid, hyphae 1–3  $\mu$ m wide. Prothallus white, up to 2 mm wide, fibrous-like. Broder line dark brownish, ca. 0.2 mm wide. Pseudoisidioid outgrowths present, usually restricted to the

Species	Known Distribution	References
C. albida (Fée) Frisch & G. Thor	Brazil, Costa Rica, Cuba, Ecuador, India, Peru, Venezuela	Aptroot et al. (2009), Frisch & Thor (2010), Behera et al. (2021), In this paper
C. athertoniensis Frisch & G. Thor	Australia	Frisch & Thor (2010)
C. bella Frisch & G. Thor	Tanzania	Frisch & Thor (2010)
<i>C. biseptata</i> (Aptroot & Wolseley) Frisch & G. Thor	New Caledonia, Thailand	Frisch & Thor (2010), Aptroot & John (2015)
C. brevispora Frisch & G. Thor	Brazil	Frisch & Thor (2010)
C. citrina Frisch & G. Thor	Ivory Coast	Frisch & Thor (2010)
C. corticorygmoides Aptroot & M.Cáceres	Brazil	Cáceres & Aptroot (2016)
C. divaricatica Aptroot & Sipman	Mexico	Aptroot et al. (2014)
C. lichexanthonica A.A. Menezes, M. Cáceres & Aptroot	Brazil	Menezes et al. (2013)
C. mycelioides (Vain.) Frisch & G. Thor	Australia, Colombia, Fiji Islands, Indonesia, Japan, Philippines, Vietnam	Aptroot & Sipman (2006), Aptroot et al. (2009), Frisch & Thor (2010), Lucking et al. 2021
C. olivacea Frisch & G. Thor	Brazil	Frisch & Thor (2010)
C. palaeotropica Frisch & G. Thor	Australia, Fiji Islands, India, Indonesia, Ivory Coast, Japan, La Réunion, Malaysia, New Zealand, Sri Lanka, Tanzania, Uganda	Frisch & Thor (2010), Frisch et al. (2015), Paukov et al. (2017), Jagadeesh Ram & Sinha (2017), In this paper
C. polillensis (Vain.) Frisch & G. Thor	Philippines, Sri Lanka	Frisch & Thor (2010), Weerakoon et al. (2016)
C. streimannii Sipman	Indonesia, Vanuatu	Sipman (2018)
C. submuriformis A.A. Menezes, M. Cáceres & Aptroot	Brazil	Menezes et al. (2013)
C. vandenboomii Frisch & G. Thor	La Réunion, Uganda	Frisch & Thor (2010), Frisch et al. (2015)

#### **Table 1:** The known distribution of the species of *Crypthonia* in the world.

peripheral parts of the thallus, short, globular to coralloid type with projecting hyphae. Photobiont cells  $5-12 \times 3-8$  µm. Ascomata white, sometimes with light rose tinge, raised, irregularly shaped, and simple to branched, up to 1.5 mm wide. Excipulum indistinct. Epithecium 5-10 µm thick. Hymenium 30–70 µm high, with brownish granules and small calcium oxalate crystals, I–. Paraphysoids highly branched and anastomosing, 1-2 µm wide. Hypothecium 40–70 µm high. Asci *Arthonia*-type, 8-spored,  $35-50 \times 12-17$  µm. Ascospores hyaline, 1-septate,  $8-14 \times 3-4.5$  µm. Pycnidia not seen.

*Chemistry*: Thallus K-, C-, P+ yellow, UV-; psoromic acid detected in TLC.

*Remarks: Crypthonia palaeotropica* is a fertile species having pseudosidia. The pseudoisidia are mainly restricted to the

peripheral regions of the thallus where ascomata are absent. So far, only four species of *Crypthonia* are reported with psesudoisidioid outgrowths, viz., *C. albida*, *C. mycelioides* (Vain.) Frisch & G. Thor, *C. palaeotropica*, and *C. streimannii* Sipman. Among these the presence of ascomata were reported only in *C. palaeotropica* and *C. streimannii*, and the latter species differs from the former species by its chemistry (containing terpenoids instead of psoromic acid) (*fide* Sipman 2018). In India, *C. palaeotropica* is known from the Andaman Islands and West Bengal (Jagadeesh Ram & Sinha 2017). The current report indicates its extended distribution to the Western Ghats biodiversity hotspot.

Specimen examined: India, Karnataka, Shimoga dist., Sagara taulk, Holebagilu, elev. 623 m a.s.l., on bark, 16 March 2001, S. Nayaka 01-222770 (LWG).

# NOTES ON THE ECOLOGY AND DISTRIBUTION OF *CRYPTHONIA*

The maximum number of species of genus Crypthonia are known from Brazil with 6 spp. (Table 1). The most common and widely distributed species is C. palaeotropica and so far, known from Australia, Fiji Islands, India, Indonesia, Ivory Coast, Japan, La Réunion, Malaysia, New Zealand, Sri Lanka, Tanzania, and Uganda. During the studies of epiphytic lichen communities of Bwindi Impenetrable National Park in African Montane Rain Forests of southwest Uganda, Frisch et al. (2015) recorded the 59 individual occurrences of C. paleotropica. Frisch & Thor (2010) noted that the occurrences of C. albida in Vanuatu and Vietnam (Aptroot et al. 2009) should be checked against C. palaeotropica. The occurrence of C. mycelioides in Vietnam (Aptroot & Sipman 2006; as Chiodecton mycelioides) is neither commented nor included in Frisch & Thor (2010). However, Joshi et al. (2018) included the species in the key to the Arthoniaceae species from Vietnam.

According to the current distributional records, the genus Crypthonia is strictly restricted to the tropics (Fig. 2). Further, most of its species are rare and known only from the type locality. The species of Crypthonia require high moisture as they are typically found in shady microhabitats and on substrates with increased water holding capacity (Frisch & Thor 2010). In India, the species are known from the tropical rain forests of Andaman Islands, Darjeeling hills, and Western Ghats. They were found growing on the decaying bark at the base of old trees along with the mosses. Frisch & Thor (2010) also noted that the Crypthonia species occurs in habitats shaded from direct rainfall, growing on soft and slightly decaying bark or over a thin layer of corticolous bryophytes or detritus that had been accumulated on the stem. However, C. mycelioides from the Philippines have been collected from rock, but no further information about the habitat is available. The two specimens of C. albida from Costa Rica were found to be growing on the living leaves (Frisch & Thor 2010).

# WORLD KEY TO THE SPECIES OF CRYPTHONIA

#### (Modified from Frisch & Thor 2010)

1a Thallus with psesudoisidioid outgrowths2		
1b Thallus without psesudoisidioid outgrowths5		
2a Thallus with norstictic acid		
3a Hypothallus olivaceous-green, ascomata and ascospores absent		

ascomata and ascospores present 4
4a Thallus with psoromic acid <i>C. palaeotropica</i> 4b Thallus with terpenoids <i>C. streimannii</i>
5a Ascospores muriformC. submuriformis5b Ascospores transversely septate66a Thallus UV+ yellow, lichexanthone present6
C. lichexanthonica
6b Thallus UV–/white, lichexanthone absent
7aAscospores up to 3-septate87b Ascospores more than 3-septate15
8a Ascospores 1-septate
9a Thallus with rugulosin, hypothallus lemon yellow <i>C. citrina</i> 9b Thallus without rugulosin, hypothallus olive-brown or 10
olive-brown C. polillensis
10b Thallus with psoromic acid, hypothallus variable 11
11a Hypothallus dark green, ascospores 7.5–10 μm long C. brevispora
11b Hypothallus olive-brown, ascospores 11–14 μm longC. vandenboomin
12a Thallus with rugulosin
13a Thallus with gyrophoric acid
13b Thallus with psoromic acid 14
14a Ascospores 10–16 μm long, widest in the upper half 
14b Ascospores 16–19 μm long, widest in the middle
15a Thallus without lichen substances, ascospores 3-5-septate C. corticorygmoides 15b Thallus with divaricatic and usnic acids, ascospores 5-9-septate
A

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